

Diversity and population structure of Criollo Lechero Tropical and Romosinuano cattle breeds in Mexico

Diversidad y estructura poblacional de las razas bovinas Criollo Lechero Tropical y Romosinuano en México

Diversidade e estrutura populacional das raças de gado Criollo Lechero Tropical e Romosinuano no México

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Abstract

Background: The assessment of genetic diversity, population structure, as well as within and across breed relationships using DNA markers is essential to develop conservation and genetic improvement programs. **Objective:** To assess the genetic diversity and population structure of Criollo Lechero Tropical (CLT) and Romosinuano (ROMO) cattle breeds in Mexico and compare them with other Criollo and Spanish breeds. **Methods:** The study included CLT and ROMO genotypes from the 65k Axiom©BovMDv3 array. Genomic data of Criollo and Spanish breeds were retrieved from a repository. Genetic diversity across breeds was evaluated by pairwise Wright's F_{ST} , principal components, and structure analysis. **Results:** Observed and expected heterozygosity, inbreeding coefficient, and effective population size (*Ne*) were 0.31, 0.30, -0.018 and 56 in CLT, and 0.32, 0.31, -0.023 and 99 in ROMO, respectively. Wright's F_{ST} indicated that both breeds are related to American Criollo and some Spanish cattle. ROMO cattle showed more significant Spanish ancestry than CLT. **Conclusions:** This study presents novel evidence indicating that, although the *Ne* of these breeds is small, they are still diverse and can be used as genetic reservoirs in conservation programs.

Keywords: *cattle*; *Criollo Lechero Tropical*; *genetic diversity*; *DNA markers*; *effective population size*; *inbreeding*; *population structure*; *Romosinuano*.

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Resumen

Antecedentes: La evaluación de la diversidad genética, de la estructura de la población y de las relaciones dentro y entre razas utilizando marcadores de ADN es esencial para el desarrollo de programas de conservación y mejora genética. **Objetivo:** Evaluar la diversidad genética y la estructura poblacional de las razas bovinas Criollo Lechero Tropical (CLT) y Romosinuano (ROMO) de México y compararlas con otras razas criollas y españolas. **Métodos:** El estudio incluyó genotipos CLT y ROMO de la matriz Axiom©BovMDv3 de 65k. Los datos genómicos de razas criollas y españolas fueron obtenidos de un repositorio. La diversidad genética entre razas se evaluó mediante el F_{ST} de Wright para pares de razas, componentes principales, y análisis de estructura. **Resultados:** La heterocigosidad observada y esperada, el coeficiente de endogamia y el tamaño efectivo de la población (*Ne*) fueron 0,31, 0,30, -0,018 y 56 en CLT, y 0,32, 0,31, -0,023 y 99 en ROMO, respectivamente. El F_{ST} de Wright indicó que ambas razas están relacionadas con el criollo americano y algunos bovinos españoles. El ganado ROMO muestra una ascendencia española más significativa que el CLT. **Conclusiones:** El estudio presenta evidencia novedosa que indica que aunque el *Ne* de estas razas es pequeño, todavía son diversas y pueden ser utilizadas como reservorios genéticos en programas de conservación.

Palabras clave: bovinos; Criollo Lechero Tropical; diversidad genética; endogamia; estructura de la población; marcadores de ADN; Romosinuano; tamaño efectivo de la población.

Resumo

Antecedentes: A avaliação da diversidade genética, da estrutura populacional e das relações dentro e de todas as raças usando marcadores de DNA é essencial para o desenvolvimento de programas de conservação e melhoramento genético. Objetivo: Avaliar a diversidade genética e a estrutura populacional das raças Criollo Lechero Tropical (CLT) e Romosinuano (ROMO) do México e compará-las com outras raças crioulas e espanholas. Métodos: O estudo incluiu genótipos CLT e ROMO da matriz Axiom©BovMDv3 de 65k. Dados genômicos de raças crioulas e espanholas foram recuperados de um repositório. A diversidade genética entre as raças foi avaliada pelo F_{ST} de Wright, principais componentes e análise de estrutura. Resultados: Heterozisia observada e esperada, coeficiente de endogamia e tamanho populacional efetivo (*Ne*) foram 0,31, 0,30, -0,018 e 56 na CLT, e 0,32, 0,31, -0,023 e 99 em ROMO, respectivamente. A F_{ST} de Wright indicou que ambas as raças estão relacionadas com o crioulo americano e alguns bovinos espanhóis. O gado ROMO mostra ascendência espanhola mais significativa do que a CLT. Conclusões: O estudo apresenta novas evidências indicando que, embora *Ne* dessas raças seja pequeno, elas ainda são diversas e podem ser utilizadas como reservatórios genéticos em programas de conservação.

Palavras-chave: Criollo Lechero Tropical; diversidade genética; endogamia; estrutura populacional; gado; marcadores de DNA; Romosinuano; tamanho populacional efetivo.

Romosinuano (ROMO) cattle is a Colombian

Criollo breed introduced to Mexico by an

absorption process of individuals and semen of

ROMO bulls from Florida, USA, and Turrialba,

Costa Rica (De Alba, 2011). ROMO cattle

The assessment of genetic diversity, population structure, and within and across breeds relationships is essential for the development of conservation and genetic improvement programs of cattle populations (Martínez et al., 2012). The genomic data of the populations, export, divergence, and admixture in domestic cattle can be precisely established (Decker et al., 2014a). Genetic markers have been used worldwide to assess genetic variation between cattle breeds in regard to their geographical origin. The SNP markers have been used to develop Chip SNP, which is a highly informative and density-based microarray technology (Matukumalli et al., 2009). The SNPs have been advantageous for assessing genetic diversity and estimate phylogenetic relationships (Decker et al., 2014a).

Criollo cattle breeds from America have their ancestral sources in Spanish breeds from the New World colonization. It is not known whether these breeds originated from one or more cattle populations (Martínez et al., 2012). The Criollo Lechero Tropical (CLT) cattle breed was developed in Nicaragua under geographic isolation of the original Spanish cattle that underwent later selection for milk production (De Alba, 2011). In Mexico, a CLT nucleus herd was established in the mid-20th century. Purebred CLT imported from Nicaragua and some bulls from Centro Agronómico Tropical de Investigación y Enseñanza (CATIE, Turrialba, Costa Rica) conformed the Mexican nucleus (Rosendo-Ponce and Becerril-Pérez, 2015). The CLT is currently widespread in tropical Mexico given it is a tropically adapted Bos taurus dairy breed. Due to its small population (less than 1000 cows), CLT is currently considered an endangered breed (FAO, 2013; Rosendo et al., 2018).

and tolerance to high temperature and relative humidity (Scharf *et al.*, 2010).

Criollo breeds evolved under different climatic conditions compared with their European ancestry, so they have probably fixed or changed allele frequencies, differing from those of their ancestors due to adaptation and artificial selection for improved performance under these conditions. In Mexico, genetic diversity of CLT and ROMO has been evaluated through pedigree analysis (Rosendo et al., 2018; Núñez-Domínguez et al., 2020). However, within and across breed genetic diversity and structure of CLT and ROMO have not been evaluated using genomic approaches. Currently, the use of variable density SNP microarrays has been implemented to evaluate genetic diversity and differentiation between breeds to provide an efficient genome-wide perspective of genetic flow or isolation in specific populations (Mastrangelo et al., 2014; Savaranan et al., 2021).

Therefore, this study was aimed to assess genetic diversity and structure of CLT and ROMO cattle breeds in Mexico and compare them with other Criollo ancestor breed populations using a medium-density SNP array.

Materials and Methods

Source of data

All CLT and ROMO animals included in this study belong to herds affiliated to Asociación Mexicana de Criadores de Ganado Romosinuano *v Lechero Tropical*, and are being used in a genetic improvement program by the breeders. Seventyfive animals were randomly sampled from each breed. Hair follicles were collected from each animal. Samples were genotyped using the Axiom[©] BovMDv3 array consisting of 63,791 SNPs (Affymetrix, Santa Clara, CA, USA). Monomorphic SNPs were excluded, resulting in 55,760 SNPs for CLT and 55,579 for ROMO. Genotypic data from an Illumina BovineSNP50 BeadChip array with 1,539 animals of 127 breeds were retrieved from a public repository (Decker et al., 2014b). The SNPs had been filtered by the authors based on a calling rate of 0.9, and monomorphic SNPs were excluded, remaining 43,043 SNPs available. To properly compare genotypic data from the present study with those of the public repository, we used a similar SNP edition to the one used by Decker et al. (2014b), which ignores the use of linkage disequilibrium. Only data from Holstein, Jersey, Angus, and Brown Swiss animals were included as European cattle breeds. Eleven native Spanish breeds (Berrenda Negro, Berrenda Colorado, Negra Andaluza, Cárdena Andaluza, Toro de Lidia, Pirenaica, Mostrenca, Terrana, Menorquina, Morucha, Retinta), five Criollo breeds (Texas Longhorn, Colombian Romosinuano, Corriente, Senepol, Florida Cracker), and three breeds of Asian indicine cattle (Guzerat, Sahiwal, Gyr) were selected for further analysis. Genotypic data were merged by SNP's name and location with CLT and ROMO data using PLINK v1.9 software (Purcell et al., 2007). Finally, a total of 18,192 SNPs were shared between the panels.

Within-breed genetic diversity

Observed (*Ho*) and expected (*He*) heterozygosity and inbreeding coefficient (F_{IS}) were calculated for the CLT and ROMO samples using the PLINK software (Purcell *et al.*, 2007). Trends of the effective population size (*Ne*) were calculated from linkage disequilibrium throughout the CLT and ROMO genomes using the SNeP V1.1 software (Barbato *et al.*, 2015).

Across-breed genetic diversity

Pairwise F_{ST} between CLT, ROMO, and Criollo, Spanish, and indicine breeds were calculated using the PLINK software (Purcell *et al.*, 2007). Principal components (PC) were calculated using the merged SNP dataset considering only autosomal SNPs. The genomic relationship matrix was calculated using the GCTA software (Yang *et al.*, 2011). The R package OmicKriging (Im *et al.*, 2016) was used to access the matrix information and subsequently estimate PC using the RSpectra package of R (Qiu *et al.*, 2019), and the top two PC were plotted against each other considering CLT, ROMO, and the five Criollo, eleven Spanish, and three indicine breeds. Using the SRUCTURE v2.3.4 software (Pritchard *et al.*, 2000), population structure analyses were performed with an initial Burn-in of 25,000, and 25,000 additional Markov Chain Monte Carlo (MCMC) runs. The dataset was processed for five iterative cycles, each with constant Burn-in and MCMC. The four European, eleven native Spanish, five Criollo, and three breeds of Asian indicine cattle were included to consider the possible subpopulations of ancestry in CLT and ROMO. K-values from 2 to 13 were tested with the STRUCTURE harvester software (Earl and vonHoldt, 2012) to infer the best fit Delta K-value between populations using the Evanno's method (Evanno *et al.*, 2005).

Results

Within-breed genetic diversity

The Ho and He, average F_{IS} , and Ne were calculated (Table 1). ROMO showed greater difference between Ho and He compared to CLT cattle. The Ne trend across the last 100 generations is shown in Figure 1. The Ne for both breeds has been decreasing over time. The generation intervals of ROMO and CLT breeds were 6.7 (Núñez-Domínguez et al., 2020) and 6.9 years (Rosendo et al., 2018), respectively. The first trips carrying cattle from Spain to America were reported in 1524 (Ajmone-Marsan et al., 2010). In 70 generations, the Ne was reduced from 317 to 56 in CLT, and from 394 to 99 in ROMO. None of these breeds had a Ne value less than the lower limit (<50) recommended for conservation of genetic resources (FAO, 1998).

Table 1. Observed (*Ho*) and expected (*He*) heterozygosity, inbreeding coefficient (F_{IS}), and effective population size (*Ne*) of Criollo Lechero Tropical (CLT) and Romosinuano cattle from Mexico.

| Breed | Но | Не | F _{IS} | Ne |
|-------------|-------|-------|-----------------|----|
| CLT | 0.310 | 0.304 | -0.018 | 56 |
| Romosinuano | 0.326 | 0.318 | -0.023 | 99 |

Across-breed genetic diversity

Wright's F_{ST} values calculated between CLT, ROMO, and established cattle breeds are shown

in Table 2. The smallest Wright's F_{ST} value among ROMO with a Criollo breed (omitting the comparison with Colombian Romosinuano) was with Texas Longhorn, while the lowest values compared to Spanish breeds were with Andalusian breeds. The same results were observed with CLT. The highest F_{ST} values were with Bos indicus breeds for both CLT and ROMO. The F_{ST} values between CLT and ROMO with Bos indicus breeds were 0.25 and 0.23, respectively.

The main two PC were calculated, and animal samples were graphically represented (Figure 2). Bos taurus and Bos indicus species showed the largest difference in principal components. Criollo and Spanish cattle breeds showed a slight separation between clusters. CLT and ROMO were clustered with the Criollo breeds; however, these breeds exhibited large spread as PC points.

Genetic structure analyses are shown in Figure 3. The best fit given all the populations was determined as K=10, indicating that both CLT and ROMO show different structure compared to the other breeds in the study. However, considering the K-value of 4, ROMO and CLT shared 80 and 81%, respectively, of a common ancestral Criollo and Spanish subpopulation breeds, which shows the relationship between Criollo and Spanish cattle, grouping in the same cluster as observed in the PC analysis. The largest ancestral subpopulation that shows ROMO in Mexico is shared with Colombian Romosinuano. This subpopulation represents, on average, 75% of ROMO genes, which at the same time, this subpopulation represents on average 7% in the Spanish breeds. The other ancestral subpopulations in the ROMO population represent 10% of the subpopulation shared mainly with CLT; 2% of the subpopulation shared with Criollo cattle such as Texas Longhorn and Corriente; and 8.7% belongs mainly to subpopulations of Bos indicus ancestors. CLT cattle shows a subpopulation that covers 89% of its ancestry. This ancestral subpopulation is not shared with high percentages in other bovine breeds. Spanish breeds such as Cárdena Andaluza, Negra Andaluza, and Berrenda en Colorado show a slight proportion of this ancestral subpopulation (3% on average).

| Table 2. Pairwis | e F_{ST} betwee | een Criollo Lechero |
|-------------------|-------------------|----------------------|
| Tropical (CLT) an | d Romosinua | no breeds with other |
| cattle breeds. | | |

| Breed | Romosinuano | CLT |
|-----------------------|-------------|--------|
| Texas Longhorn | 0.0760 | 0.0791 |
| Colombian Romosinuano | 0.0769 | 0.1006 |
| Corriente | 0.0795 | 0.0825 |
| Senepol | 0.0822 | 0.0854 |
| Florida Cracker | 0.1191 | 0.1188 |
| Berrenda Negro | 0.0698 | 0.0685 |
| Berrenda Colorado | 0.0757 | 0.0768 |
| Negra Andaluza | 0.0800 | 0.0802 |
| Cárdena Andaluza | 0.0821 | 0.0828 |
| Toro de Lidia | 0.0854 | 0.0850 |
| Pirenaica | 0.0860 | 0.0860 |
| Mostrenca | 0.0882 | 0.0876 |
| Terrana | 0.0971 | 0.0960 |
| Menorquina | 0.1027 | 0.1023 |
| Morucha | 0.1512 | 0.1513 |
| Retinta | 0.1612 | 0.1611 |
| Angus | 0.1204 | 0.1203 |
| Brown Swiss | 0.1343 | 0.1369 |
| Holstein | 0.1082 | 0.1035 |
| Jersey | 0.1513 | 0.1447 |
| Guzerat | 0.2083 | 0.2275 |
| Sahiwal | 0.2394 | 0.2584 |
| Gir | 0.2417 | 0.2603 |

Discussion

The Ho, He, and F_{IS} values have been used to reference genetic diversity of small and indigenous populations. The Ho values were higher than He in both breeds of the present study. Similarly, *Ho* values were higher than *He* in Tharparkar cattle (Saravanan et al., 2020). Also, the Korea Brown Hanwoo, Brindle Hanwoo, and Jeju Black native cattle showed great genetic diversity, having *Ho* greater than *He* (Sharma *et al.*, 2016). Unfortunately, the sample of Colombian ROMO in the Decker's et al. (2014) paper was too small to make meaningful comparisons with the results of the present study.

The negative F_{IS} values in the present study (-0.018 and -0.023 for CLT and ROMO, respectively) correspond to inbreeding levels

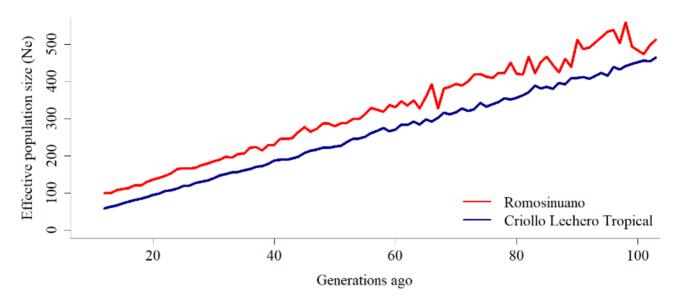


Figure 1. Estimated effective population size trend from the last 100 generations.

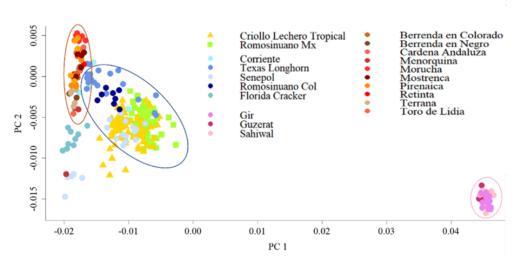


Figure 2. Principal components plot of Criollo Lechero Tropical, Mexican Romosinuano, American Criollo, Spain native and indicine cattle breeds.

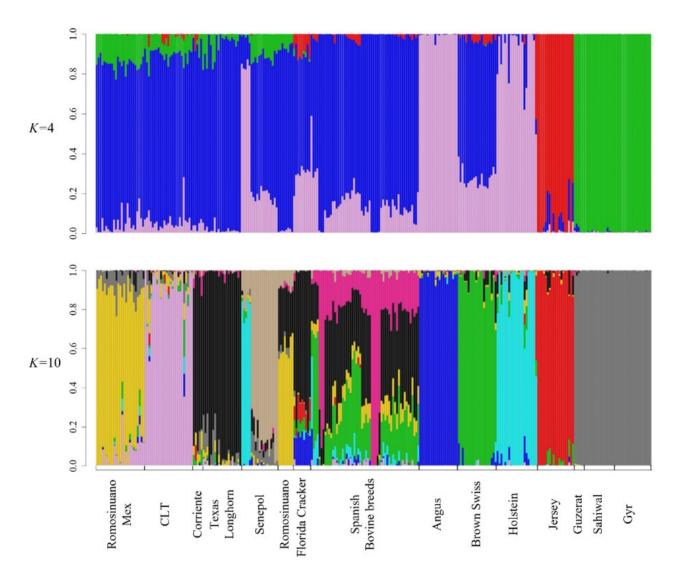


Figure 3. Structure analysis with K=4 and K=10 (best fit) in Criollo Lechero Tropical, Romosinuano, and cattle reference populations.

lower than average homozygosity of the populations (0.310 for CLT and 0.326 for ROMO). Different results were obtained by Eusebi *et al.* (2017) who reported average F_{IS} values of 0.06 and 0.01 for the same repository data from the Spanish and Criollo breeds used in the present study. Likewise, Rosendo *et al.* (2018) and Núñez-Domínguez *et al.* (2020), based on a pedigree information analysis, estimated 0.01 and 0.025 inbreeding coefficients for CLT and ROMO, respectively. Using SNP, the parameters calculated in the present study showed higher genetic diversity within breeds than the pedigree analysis.

Therefore, within-breed genetic diversity studies are necessary for Criollo and local breeds to be used in a productive and economic environment and to avoid inbreeding problems (Meuwissen, 2009).

In general, the differences with Spanish and Criollo breeds are categorized as 'moderate differentiation' for both breeds, while 'great differentiation' was observed with indicine breeds (Weir and Cockerham, 1984). The lowest Wright's F_{ST} values among Mexican ROMO, CLT, and the Spanish breeds agree with reports mentioning that cattle arrival to the New World occurred in the

second and later trips of Christopher Columbus. Cattle left Seville and nearby ports, so Andalusian cattle would be the most likely ancestors of Criollo breeds (Ajmone-Marsan *et al.*, 2010).

Gautier *et al.* (2010) reported F_{ST} values between European cattle and *Bos indicus* breeds between 0.29 and 0.38. This result is consistent with Decker *et al.* (2014a) and Martínez *et al.* (2012) who mention that Criollo breeds have indicine ancestry, which occurred after the arrival of Spanish cattle to the New World. Miretti *et al.* (2004) suggest that Zebu's influence in Criollo cattle may be due to imports of *Bos indicus* from Africa to improve adaptability to tropical conditions during the colonial period.

The long-term genetic variation maintained in the population is proportional to the effective population size (FAO, 1998). The Ne from genealogical records was calculated for CLT and ROMO in México (Rosendo et al., 2018; Núñez-Domínguez et al., 2020). One way to estimate Ne is by using pedigree; however, these estimates depend on the integrity of genealogical records (Barbato et al., 2015). According to the Ne trends estimated in the present study the Ne value published by pedigree analysis (Rosendo et al., 2018) was reached thirteen generations earlier, while the value has not yet been reached in ROMO (Rosendo et al., 2018). Hidalgo et al. (2021) estimated a Ne of 72 in generation 1 from a genomic sample of the same ROMO population in Mexico. FAO (2013) suggested a minimum Ne of 50 per generation to maintain genetic diversity. The estimated Ne in our study was above 50.

Estimates of F and Ne values in CLT from the present study (-0.018 and 56, respectively) differ from those of Rosendo *et al.* (2018) using pedigree analyses (0.043 and 68.1). Similarly, estimates of F and Ne in ROMO from the present study (-0.023 and 99, respectively) differ from those of Núñez-Domínguez *et al.* (2020) using pedigree analyses (0.026 and 45, respectively). These differences are expected since estimates from pedigree analyses are based on probabilities of common genes among relatives, while genomic analyses rely on common SNP markers; therefore, the latter procedure is more accurate. Conservation strategies for CLT and ROMO should consider estimates from genomic analyses to support better decision making.

CLT and ROMO remained close to Criollo breeds in the PC plot, mainly with Texas Longhorn, Corriente, and Senepol, confirming the slight differentiation from native Spanish and wholly differentiated from *Bos indicus*-like observed in the Wright's F_{ST} comparison. Strucken *et al.* (2021) mentioned that *Bos indicus* breeds show low across-breed genetic diversity among breeds because the *Ne* of Indian indigenous breeds have been large since domestication. Contrarily, criollo breeds were developed with a small *Ne* since the arrival of the Spanish people to America; therefore, criollo breeds show wide across-breed genetic diversity.

The PC analysis considers a fraction of the variance included in the genotyped animals, which explains this variability in terms of a minimum of principal components (Figure 2). Pritchard *et al.* (2000) suggested STRUCTURE as an exploratory tool when inspecting results from a range of values of K. A delta K-value of 10 was the best at separating cattle subpopulations, being sensitive to the number of breeds and population size. A delta K-value of 4 clearly separates (Figure 3) four subpopulation groups (*Bos taurus, Bos indicus,* Criollo, and Spanish cattle), in agreement with the clusters observed in the PC analysis plot (Figure 2).

The results of the structure analysis support Wright's F_{ST} coefficients. ROMO cattle has a low percentage of indicine origin ancestry present in some Criollo populations (Decker *et al.*, 2009; Martínez *et al.*, 2012; Decker *et al.*, 2014a). CLT cattle only presented 2% of *Bos indicus* ancestors. Using only 19 microsatellite markers and calculating the contributions by the likelihood estimation of admixture proportions with the LEADMIX software, Martínez *et al.* (2012) found that Iberian breeds significantly contributed to Criollo cattle (between 70 and 80%). With the same number of microsatellite markers and a high K-level, a set of Criollo bovine breeds did not show clear admixture evidence with any of the other breeds evaluated (Ginja *et al.*, 2019). There was a lower proportion of ancestry in other subpopulations of European breeds due to origin differences of British, continental, and Spanish cattle (Decker *et al.*, 2009). Our results strongly support the idea that Criollo breeds derived their own genetic identity and deserve to be adequately managed and conserved.

ROMO and CLT in America have originated different genetic structures between European, Spanish. and Criollo breeds. Exportation. admixture, and breed formation greatly impact variation between breeds (Decker et al., 2014a). Breed diversity is helpful because it provides alternatives when commercial breeds have problems due to genetic drift or if changes in the production sector require poorly developed traits in the commercial breeds (Weir and Cockerham, 1984). For the assessed breeds, our results suggest enough diversity as a genetic reservoir considering their essential advantages in tropical environments (Hammond et al., 1996; Johnson et al., 2012; Becerril-Pérez et al., 2020) and preserving their productivity in these conditions (Rosendo-Ponce and Becerril-Pérez, 2015; Riley et al., 2015).

In conclusion, although CLT and ROMO cattle represent small populations, within-breed genetic diversity parameters confirm their degree of heterozygosity. CLT and ROMO diverge from ancestor Criollo populations suggesting that they developed their own genetic pool during adaptation to the challenging environment. Given that CLT and ROMO breeds are tropically adapted *Bos taurus*, they have become a valuable genetic resource that justifies structured and constant conservation and utilization programs to preserve their genetic diversity.

Declarations

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Conflicts of interest

The authors declare they have no conflicts of interest regarding the work presented in this report.

Author contributions

All authors conceived and validated the study, contributed to methods and to writing the paper; data curation and formal analysis, Ricardo E. Martínez-Rocha, Gaspar Manuel Parra-Bracamonte and José G. García-Muñiz; writing-review and editing, Rodolfo Ramírez-Valverde, Rafael Núñez-Domínguez and José G. García-Muñiz; supervision, Rodolfo Ramírez-Valverde and Gaspar Manuel Parra-Bracamonte; management to get support from CONARGEN, Rafael Núñez-Domínguez. All authors read and agreed to the published version of the manuscript.

Use of artificial intelligence (AI)

No AI or AI-assisted technologies were used during the preparation of this work.

References

Ajmone-MarsanP,GarciaF,LenstraJA.Ontheorigin of cattle: how aurochs became cattle and colonized the world. Evol Anthropol 2010; 19(4):148–157. https://doi.org/10.1002/evan20267

Barbato M, Orozco-terWengel P, Tapio M, Bruford MW. SNeP: a tool to estimate trends in recent effective population size trajectories using genome-wide SNP data. Front Genet 2015; 6:109. https://doi.org/10.3389/fgene.2015.00109 Becerril-Pérez CM, Álvarez-Cepeda AA, Rosendo-Ponce A, Alonso-Morales RA. Kappacasein genotyping in tropical milking Criollo and its association to milk production and composition. Trop Anim Health Prod 2020; 52(6):3885–3888. https://doi.org/10.1007/s11250-020-02317-3

Carroll JA, Burdick NC, Reutera RR, Chase CCJr, Spiers DE, Arthington JD, Coleman SW. Differential acute phase immune responses by Angus and Romosinuano steers following an endotoxin challenge. Domest Anim Endocrinol 2011; 41(4):163–173. https://doi.org/10.1016/j. domaniend.2011.06.002

Carroll JA, Burdick NC, Chase CCJr, Coleman SW, Spiers DE. Influence of environmental temperature on the physiological, endocrine, and immune responses in livestock exposed to a provocative immune challenge. Domest Anim Endocrinol 2012; 43(2):146–153. https://doi.org/10.1016/j.domaniend.2011.12.008

De Alba JM. El libro de los bovinos criollos en América. Mexico DF: Mundi Prensa Mexico; 2011.

Decker JE, Mckay SD, Rolf MM, Kim J, Alcalá AM, Sonstegard TS, Hanotte O, Seabury CM, Praharani L, Go A, Liu W, Lei C, Reecy JM, Saif-ur-rehman M, Schnabel RD, Taylor JF. Worldwide patterns of ancestry, divergence, and admixture in domesticated cattle, PLOS Genet 2014a; 10(3):e1004254. https://doi.org/10.1371/journal.pgen.1004254

Decker JE, McKay SD, Rolf MM, Kim J, Alcalá AM, Sonstegard TS, Taylor JF. Data from: Worldwide patterns of ancestry, divergence, and admixture in domesticated cattle. Dryad Digital Repository 2014b. <u>https://doi.org/10.5061/dryad.th092</u>

Decker JE, Pires JC, Conant GC, McKay SD, Heaton MP, Chen K, Cooper A, Vilkki J, Seabury CM, Caetano AR, Johnson GS, Brenneman RA, Hanotte O, Eggert LS, Wiener P, Kim J-J, Kim KS, Sonstegard TS, Van Tassell CP, Neibergs HL, McEwan JC, Brauning R, Coutinho LL, Babar ME, Wilson GA, McClure MC, Rolf MM, Kim J, Schnabel RD, Taylor JF. Resolving the evolution of extant and extinct ruminants with high-throughput phylogenomics. Proc Natl Acad Sci USA 2009; 106(44):18644 LP–18649. <u>https://doi.org/10.1073/pnas.0904691106</u>

Earl DA. vonHoldt BM. **STRUCTURE** HARVESTER: а website and program visualizing **STRUCTURE** output for and implementing the Evanno method. Conserv Genet Resour 2012; 4(2):359-361. https://doi.org/10.1007/s12686-011-9548-7

Eusebi PG, Cortés O, Dunner S, Cañón J. Genomic diversity and population structure of Mexican and Spanish bovine Lidia breed. Anim Genet 2017; 48(6):682–685. <u>https://doi.org/https://doi.org/10.1111/age.12618</u>

Evanno G, Regnaut S, Goudet J. Detecting the number of clusters of individuals using the software structure: a simulation study. Mol Ecol 2005; 14(8):2611–2620. <u>https://doi.org/10.1111/j.1365-294X.2005.02553.x</u>

Food and Agriculture Organization of the United Nations (FAO). Secondary Guidelines for Development of National Farm Animal Genetic Resources Management Plans: Management of Small Populations at Risk. Rome, Italy; 1998.

Food and Agriculture Organization of the United Nations (FAO). *In vivo* conservation of animal genetic resources. Animal Production and Health Guidelines 14. Rome Italy; 2013.

Gautier M, Laloë D, Moazami-Goudarzi K. Insights into the genetic history of French cattle from dense SNP data on 47 worldwide breeds. PLoS One 2010; 5(9):e13038. https://doi.org/10.1371/journal.pone.0013038

Ginja C, Gama LT, Cortés O, Burriel IM, Vega-Pla JL, Penedo C, Sponenberg P, Cañón J, Sanz A, Alves A, Alvarez LA, Giovambattista G, Agha S, Rogberg-Muñoz A, Lara MAC, Consortium B, Delgado JV, Martinez A. The genetic ancestry of American Creole cattle inferred from uniparental and autosomal genetic markers. Sci Rep 2019; 9(1):11486. <u>https://doi.org/10.1038/s41598-019-47636-0</u> Hammond AC, Olson TA, Chase Jr CC, Bowers EJ, Randel RD, Murphy CN, Vogt DW, Tewolde A. Heat tolerance in two tropically adapted *Bos taurus* breeds, Senepol and Romosinuano, compared with Brahman, Angus, and Hereford cattle in Florida. J Anim Sci 1996; 74(2):295–303. https://doi.org/10.2527/1996.742295x

Hidalgo J, Cesarani A, Garcia A, Sumreddee P, Larios N, Mancin E, García JG, Núñez R, Ramírez R. Genetic background and inbreeding depression in Romosinuano cattle breed in Mexico. Anim 2021; 11(2):321. https://doi.org/10.3390/ani11020321

Im HK, Wheeler HE, Michaels KA, Trubetskoy V. Package 'OmicKriging' 2016. <u>https://pbil.univ-lyon1.fr/CRAN/web/packages/OmicKriging/</u><u>OmicKriging.pdf</u>

Johnson JS, Scharf B, Weaber RL, Eichen PA, Spiers DE. Patterns of heat response and adaptation on summer pasture : A comparison of heatsensitive (Angus) and -tolerant (Romosinuano) cattle. J Therm Biol 2012; 37(4):344–350. https://doi.org/10.1016/j.jtherbio.2011.10.014

Martínez AM, Gama LT, Cañón J, Ginja C, Delgado JV, Dunner S, Landi V, Martín-Burriel I, Penedo MCT, Rodellar C, Vega-Pla JL, Acosta A, Álvarez LA, Camacho E, Cortés O, Marques JR, Martínez R, Martínez RD, Melucci L, Martínez-Velázquez G, Muñoz JE, Postiglioni A, Quiroz J, Sponenberg P, Uffo O, Villalobos A, Zambrano D, Zaragoza P. Genetic footprints of iberian cattle in America 500 years after the arrival of Columbus. PLoS One 2012; 7(11):e49066. https://doi.org/10.1371/journal.pone.0049066

Mastrangelo S, Saura M, Tolone M, Salces-Ortiz J, Di Gerlando R, Bertolini F, Portolano B. The genome-wide structure of two economically important indigenous Sicilian cattle breeds. J Anim Sci 2014; 92(11):4833–4842. https://doi.org/10.2527/jas.2014-7898

Matukumalli LK, Lawley CT, Schnabel RD, Taylor JF, Allan MF, Heaton MP, Connell JO, Moore SS, Smith TPL, Sonstegard TS, Van Tassell CP. Development and characterization of a high

Rev Colomb Cienc Pecu 2024; 37(3, Jul-Sep):123–134 https://doi.org/10.17533/udea.rccp.v37n3a2 density SNP genotyping assay for cattle. PLoS One 2009; 4(4):e5350. <u>https://doi.org/10.1371/journal.pone.0005350</u>

Meuwissen T. Genetic management of small populations: A review. Acta Agr Scand A-AN 2009; 59(2):71–79. <u>https://doi.org/10.1080/09064700903118148</u>

Miretti MM, Dunner S, Naves M, Contel EP, Ferro JA. Predominant African-derived mtDNA in Caribbean and Brazilian Creole Cattle is also found in Spanish cattle (*Bos taurus*). J Hered 2004; 95(5):450–453. https://doi.org/10.1093/jhered/esh070

Núñez-Domínguez R, Martínez-Rocha RE. Hidalgo-Moreno JA, Ramírez-Valverde R. García-Muñiz JG. Evaluation of the cattle population structure Romosinuano Mexico using pedigree analysis. Rev in Cienc Pecu 2020; 33(1):44-59. Colomb https://doi.org/10.17533/udea.rccp.v32n4a05

Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multilocus genotype data. Genetics 2000; 155(2):945–959. https://doi.org/10.1093/genetics/155.2.945

Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, Maller J, Sklar P, de Bakker PIW, Daly MJ, Sham PC. PLINK: A tool set for whole-genome association and populationbased linkage analyses. Am J Hum Genet 2007; 81(3):559–575. <u>https://doi.org/10.1086/519795</u>

Qiu Y, Mei J, Guennebaud G, Niesen J. RSpectra: solvers for large scale eigenvalue and SVD problems. R Package Version 0.15-0 2019. https://CRAN.R-project.org/package=RSpectra

Riley DG, Chase CCJr, Coleman SW, Olson TA. Evaluation of birth and weaning traits of Romosinuano calves as purebreds and crosses with Brahman and Angus. J Anim Sci 2007; 85(2): 289–298. <u>https://doi.org/10.2527/jas.2006-416</u>

Riley DG, Burke JM, Chase CC, Coleman SW. Genetic effects for reproductive performance of straightbred and crossbred Romosinuano and Angus cows in a temperate zone. Livest Sci 2015; 180(10):22–26. <u>https://doi.org/10.1016/j.</u> <u>livsci.2015.06.024</u>

Rosendo PA, Palacios JAL, Rosales MF, Torres HG, Ramírez VR, Becerril PCM. Genetic variability of Tropical Milking Criollo cattle of Mexico. Rev Colomb Cienc Pecu 2018; 31(3):196–203. https://doi.org/10.17533/udea.rccp.v31n3a04

Rosendo-Ponce A, Becerril-Pérez CM. Avance en el conocimiento del bovino Criollo Lechero Tropical en México. Ecos Rec Agrop 2015; 2(5):233–243. <u>https://era.ujat.mx/index.php/rera/</u> <u>article/view/760/643</u>

Saravanan KA, Panigrahi M, Kumar H, Parida S, Bhushan B, Gaur GK, Kumar P, Dutt T, Mishra BP, Singh RK. Genome-wide assessment of genetic diversity, linkage disequilibrium and haplotype block structure in Tharparkar cattle breed of India. Anim Biotechnol 2020; 33(2):297–311. https://doi.org/10.1080/10495398.2020.1796696

Saravanan KA, Panigrahi M, Kumar H, Parida S, Bhushan B, Gaur GK, Dutt T, Mishra BP, Singh RK. Genomic scans for selection signatures revealed candidate genes for adaptation and production traits in a variety of cattle breeds. Genomics 2021; 113(3):955–963. https://doi.org/10.1016/j.ygeno.2021.02.009 Scharf B, Carroll JA, Riley DG, Chase CCJr, Coleman SW, Keisler DH, Weaber RL, Spiers DE. Evaluation of physiological and blood serum differences in heat-tolerant (Romosinuano) and heat susceptible (Angus) *Bos taurus* cattle during controlled heat challenge. J Anim Sci 2010; 88(7):2321–2336. <u>https://doi.org/10.2527/jas.2009-2551</u>

Sharma A, Lee S-H, Lim D, Chai H-H, Choi B-H, Cho Y. A genome-wide assessment of genetic diversity and population structure of Korean native cattle breeds. BMC Genet 2016; 17:139. https://doi.org/10.1186/s12863-016-0444-8

Strucken EM, Gebrehiwot NZ, Swaminathan M, Joshi S, Al Kalaldeh M, Gibson JP. Genetic diversity and effective population sizes of thirteen Indian cattle breeds. Genet Sel Evol 2021; 53(47):1–17. <u>https://doi.org/10.1186/s12711-021-00640-3</u>

Weir BS, Cockerham CC. Estimating F-statistics for the analysis of population structure. Evolution 1984; 38(6):1358–1370. <u>https://doi.org/10.2307/2408641</u>

Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: A tool for genome-wide complex trait analysis. Am J Hum Genet 2011; 88(1):76–82. https://doi.org/10.1016/j.ajhg.2010.11.011