

PHYSICOCHEMICAL CHARACTERIZATION OF ÑUÑA BEAN (*Phaseolus vulgaris* L.) PROTEIN EXTRACT

CARACTERIZACIÓN FISIQUÍMICA DE EXTRACTO PROTEICO DEL FRIJOL ÑUÑA (*Phaseolus vulgaris* L.)

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ABSTRACT

Background: Although legume protein extracts are useful in food preparation and processing as foam stabilizers and as viscosity, palatability and nutrition enhancers, many legume proteins from South America have not been characterized extensively. One such legume is the ñuña bean (*Phaseolus vulgaris* L.), which is cooked using dry heat until the cotyledons rapidly expand with a pop. The bean is widely cultivated in the Andes, but almost unknown elsewhere. **Objective & Methods:** In this study, we characterized ten functional properties of a ñuña protein extract using standard food analysis methods. **Results:** The extract was similar to other legume protein extracts for many properties (amino acid profile, proximate analysis, yield, water absorption, color, isoelectric point, and thermogravimetric analysis). The electrophoretic analysis revealed that the sample was nearly pure phaseolin. Additionally, the ability to form foam and increase solution viscosity were comparatively low when contrasted to other extracts. **Conclusion:** These properties make ñuña protein extract useful as a nearly pure phaseolin nutrition enhancer in beverages where foaming and high viscosity are undesirable, such as in fortified beverages, drinkable yogurts, or protein supplements. The extract may also have relevance as a weight-loss supplement. Therefore, we expect that incorporating ñuña protein in processed foods would be a straightforward process.

Keywords: phaseolin, functional properties, *Phaseolus*, food additive, amino acid

RESUMEN

Antecedentes: Los extractos proteicos de leguminosas son muy utilizados en la preparación y procesamiento de alimentos como agentes estabilizadores de espuma y viscosidad, así como potenciadores de palatabilidad y nutrición. Sin embargo, muchas proteínas de leguminosas procedentes de Sudamérica no han sido caracterizadas extensamente. Una de ellas es el frijol ñuña (*Phaseolus vulgaris* L.), el cual se cocina utilizando calor seco hasta que los cotiledones se expanden rápidamente y explotan. La ñuña se cultiva ampliamente en los Andes, pero es mayormente desconocida en otras partes del mundo. **Objetivo y Métodos:** En el presente estudio, caracterizamos diez propiedades funcionales de un extracto proteico de ñuña, utilizando métodos estándares para análisis de alimentos. **Resultados:** Varias propiedades

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del extracto analizado fueron similares a las de los extractos proteicos de otras leguminosas (perfil de aminoácidos, análisis proximal, rendimiento, absorción de agua, color, punto isoeléctrico y análisis termogravimétrico). El análisis electroforético reveló que la muestra es mayormente faseolina. Además, el extracto analizado presentó baja capacidad para formar espuma e incrementar viscosidad de una solución a comparación de los otros extractos. **Conclusión:** Los resultados obtenidos indican que el extracto proteico de ñuña, que es casi faseolina pura, puede ser muy útil como potenciador nutricional de bebidas en las que la espuma y alta viscosidad son indeseadas, como es el caso de bebidas fortificadas, yogures bebibles o suplementos proteicos. El extracto podría tener relevancia como suplemento para pérdida de peso. Por lo tanto, esperamos que el uso de proteína de ñuña sea un proceso sencillo en la industria de alimentos procesados.

Palabras clave: faseolina, propiedades funcionales, *Phaseolus*, aditivo alimentario, aminoácido

INTRODUCTION

Legume protein extracts have relevant physical, nutritional, and functional properties in the food processing industry, including emulsifying, nutritional, thickening, and water absorption abilities. As a result, extensive research has been completed on characterizing nearly all aspects of common cultivars of legumes, pulses, and their extracts (1, 2). Although the characterization of legume protein has been an area of focused research for decades, extracts from legume varieties cultivated in South America have yet to be characterized thoroughly.

One cultivar of the common bean (*Phaseolus vulgaris* L.), native to the central Andes of Peru, is known as the ñuña, numia, or Andean popping bean (ÑB) (3-8). The ÑB is similar to other beans in size (0.5 to 1 g) and morphology (3, 4, 9). However, instead of being boiled before human consumption, ÑB is toasted, fried, parched, or microwaved in a similar manner to popcorn, which causes the seed coat to burst and the cotyledons to enlarge (5, 10, 11). The resulting product has an agreeable flavor similar to popcorn, toasted soy nuts, or peanuts with a texture similar to malted milk ball (10,11). Additionally, ÑB fat content (~1.3%) is lower than that of other beans, making a defatting step unnecessary during protein isolation (4, 11-13).

ÑB is widely available in its native region (8, 9, 13, 14), where it is cooked and eaten as a side dish or as a lightweight, easily transportable snack (8, 11, 13, 14). Production in Perú ranged from 1,150 to 1,831 metric tons between 2008 and 2017 (15, 16), making it available for at least the niche

market. Additionally, ÑB is slowly becoming more familiar outside of its native range. US-based and Andean nation researchers are presently taking more serious notice of the potential of ÑB (13, 14, 17-22). Research has already been conducted on the genetics of popping properties of ÑB (21, 22) and on producing cultivars that can withstand the day-length variations of a temperate climate (10, 22). At present, there are at least two consumer-oriented US providers of the bean (23, 24).

Given that ÑB is growing in familiarity worldwide but lacks extensive characterization, we analyze ten important physicochemical properties of ÑB protein extract, focusing on using this extract as a nutritional additive in processed foods.

MATERIALS AND METHODS

Ñuña beans (ÑB)

Pava variety ÑB (3, 4, 14) were obtained on the open market in Cajamarca, Perú.

Preparation of ñuña bean protein extract (ÑBPE)

ÑB were rinsed in water for 10 minutes, peeled manually, dried for 12 hours at 32 °C, and ground to an 80-mesh (0.18 mm) size. For each production lot, 50.0 g of the resulting flour was used. The flour was combined with 750 mL of distilled water. The pH was adjusted to 10-11 using concentrated NaOH (aq). The suspension was agitated at room temperature (1 h, 7,700 g, 20 °C). The pellet was discarded, and the pH of the supernatant was adjusted to 3.5 with 1N HCl (aq). The protein was precipitated for 15 minutes

at 4 °C. The suspension was centrifuged (7,700 g, 20 min, 10 °C), and the supernatant was discarded. The precipitate was rinsed with distilled water (5 mL of water per g precipitate), centrifuged again (7,700 g, 20 min, 10 °C), and the supernatant was discarded. The rinse step was repeated. The resulting protein pellet was dried using an oven (32 ± 3 °C for 14 h) or lyophilizer (Free Zone 4.5 L, LABCONCO) to produce ÑBPE. The desiccated pellet was stored at room temperature and weighed to the nearest 0.1 g. (25)

Protein extraction yield

Protein content of bean flour and protein extract were measured in duplicate using the Micro-Kjeldahl method (26) with a nitrogen-to-protein conversion factor of 5.7. The extraction yield was determined as the per-lot ratio of the mass of protein in ÑBPE to the total mass of protein in the bean flour precursor expressed as a percentage.

Foaming properties

ÑBPE (1.5 g) was combined with phosphate buffer (pH 7, 100 mL), mixed in a blender for 5 minutes, and rinsed with 10 mL of water into a graduated cylinder. Volume readings were taken 1 and 20 minutes after mixing. The before mixing volume (110 mL) was compared to the subsequent volume readings to determine foam percentage. Foaming capacity and foam stability were calculated as previously reported (27).

Water absorption properties

ÑBPE (0.5 g) and distilled water (7.5 mL) were combined in a tared tube and mixed for 1 minute. The sample was incubated at room temperature for 30 minutes, and centrifuged (1,000 g, 30 min). The supernatant was decanted, and the mass of the wet pellet was measured. Water absorption capacity (WAC) was calculated as the mass of water retained per mass of dry pellet (28).

Proximate Analysis

Moisture, ash, and fat content analyses were performed in duplicate according to standard methods (26). The protein content of ÑBPE was determined using standard Kjeldahl, Bradford, and Biuret methods. The Kjeldahl method was discussed previously. The Bradford and Biuret assays were performed according to manufacturer instructions (Sigma Aldrich ®, St. Louis; product

numbers B6916 and T1949) using bovine serum albumin standards. Bradford and Biuret analyses were performed in triplicate, and results reported as the percentage of protein in the sample. Total carbohydrate was calculated by subtracting the total content of water, protein, fat, and ash from the total mass of ÑBPE.

Color analysis

The color of ÑBPE was determined using a Minolta CR-400 colorimeter that used CIELAB color space (L a* b*) and performed in duplicate.

Isoelectric point determination

ÑBPE (0.5 g) was dissolved in 25 mL of distilled water, and pH was adjusted to 3, 4, 5, 6, 7, and 8 using 1M HCL or 1M NaOH. The suspensions were stirred for 2 h at room temperature and then centrifuged (10 min, 4,800 g, 4°C). The protein concentration of the supernate was measured by Bradford assay. Solubility was expressed as the ratio of the protein content in the supernatant over the total protein in the sample; the isoelectric point was considered to be the pH with minimum solubility.

Electrophoretic analysis of protein extract

ÑBPE was separated using sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). The sample was dissolved in a saline phosphate buffer (PBS) at a concentration of 1 mg/mL of protein, and heat-denatured with Laemmli lysis buffer for 3 min at 90 °C (29) at 2:1 sample: Laemmli buffer ratio. The sample (15 µL) was run in a Mini Protean II unit (Bio-Rad Laboratories, Hercules, CA) at 100 V for 30 min, using 4% stacking and a 10% resolving polyacrylamide gel. A wide range of unstained protein standard was used as a mass reference (Novex-Thermo Scientific). Bands in the gel were stained using Coomassie blue.

Viscosity analysis of protein extract

An aqueous 25% w/v suspension of ÑBPE was prepared and adjusted to pH 7. Viscosity at 20 °C was measured using a Brookfield viscosimeter (model RVDV – III+, USA) with a 21 cm spindle at 100 rpm.

Thermogravimetric analysis (TGA)

TGA was conducted using a Malvern TGA-Q500 analyzer (New Castle, USA). ÑBPE (6 mg) was heated from 20 °C to 1,000 °C at a rate of 20 °C/min

using a nitrogen flow rate of 90 mL/min. The loss of mass was measured as a function of temperature. The negative first derivative of the curve was taken to locate inflection points.

Amino acid analysis

A sample of protein extract was sent to a certified laboratory (Bio-Synthesis, Inc., Lewisville, Texas) for amino acid testing. $\tilde{N}BPE$ (50 mg) was hydrolyzed in 6 N HCl for 24 h at 110 °C. The product was resuspended in water and derivatized with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate. The sample was then quantized for all amino acids using an HPLC-fluorescence system. Glutamic acid and glutamine, and aspartic acid and asparagine are reported together. This method typically has a low probability of recovery of tryptophan and cysteine. Results were compared to previously published work by plotting each amino acid concentration of one sample against the same amino acid concentration of another and calculating the coefficient of determination (R^2).

RESULTS

Composition and physicochemical properties

Table 1 summarizes measured physicochemical properties of the $\tilde{N}BPE$.

Table 1. Physicochemical properties of $\tilde{N}BPE$. Appropriate units are provided, and standard deviations reported where appropriate.

Parameter	Value
Bean flour protein content	19.1%
Protein Concentration of Extract	Biuret: $91 \pm 0.6\%$ Bradford: $84.5 \pm 1.2\%$ Kjeldahl: $90.3 \pm 0.7\%$
Protein Extraction Yield	75-80%
Humidity	$3.3 \pm 0.2\%$
Fat	$3.9 \pm 0.6\%$
Ash	$2.4 \pm 0.2\%$
Carbohydrate (by difference)	$\sim 2.4\%$
Foaming Capacity	50%
Foam Stability	50% reduction in foam in 20 min
Water Absorption Capacity	3.5 g water / g protein extract
Color (L, a*, b*)	77.34, 0.22, 16.54
Viscosity (25% w/v solution)	27.5 ± 2 mPa*s

Parameter	Value
Essential Amino Acids (g/100 g extract)	Histidine 2.22
	Isoleucine 3.55
	Leucine 6.91
	Lysine 6.47
	Methionine 0.86
	Phenylalanine 4.72
	Threonine 2.86
Valine 3.96	
Conditionally Essential Amino Acids (g/100 g extract)	Arginine 4.33
	Cysteine 0.08
	Glycine 2.39
	Proline 2.76
Non-Essential Amino Acids (g/100 g extract)	Tyrosine 2.71
	Alanine 2.85
	Aspartic Acid + Asparagine 10.39
	Glutamic Acid + Glutamine 13.75
	Serine 4.59

Solubility and isoelectric point

The solubility of $\tilde{N}BPE$ is high at both low and high pH, resulting in a U-shaped solubility curve. Given that protein's solubility decreases at its isoelectric point due to the loss of favorable water-protein interactions, the isoelectric point of $\tilde{N}BPE$ can be estimated from the observed solubility minimum between pH 4 and 5 (Figure 1).

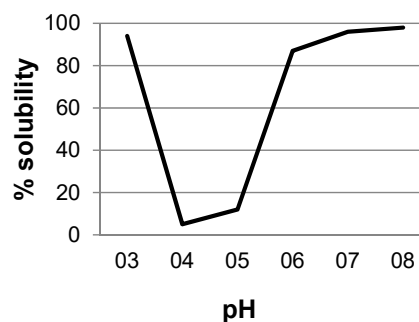


Figure 1. $\tilde{N}BPE$ solubility as a function of pH.

Electrophoretic analysis

$\tilde{N}BPE$ was separated by electrophoresis (Figure 2). The gel image shows three bright bands between 40 and 50 kDa that accounts for nearly all of the protein in the extract.

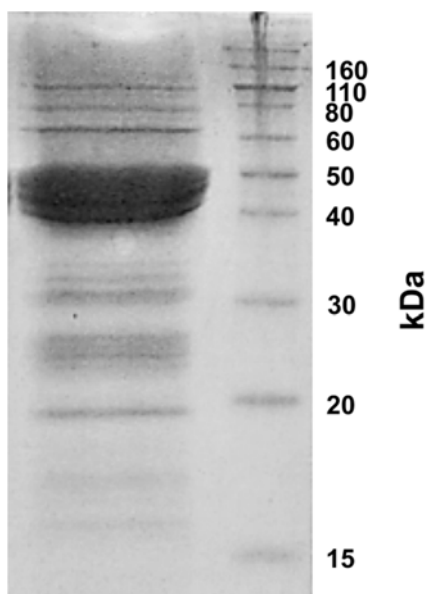


Figure 2. SDS-PAGE gel image of ÑBPE (left) and molecular weight marker (right).

Thermogravimetric analysis (TGA)

There were four inflection points on the mass curve as the heat was increased (Figure 3), indicating that there are four major steps in the sample's pyrolysis.

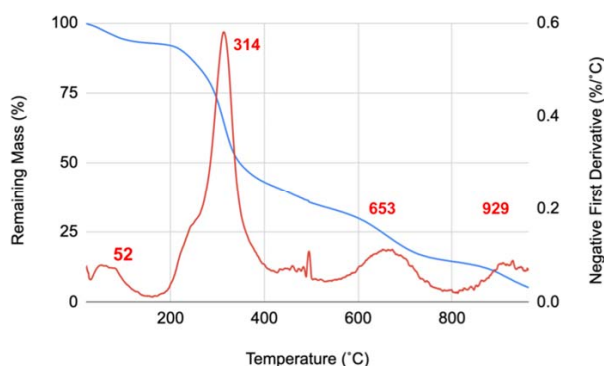


Figure 3. Thermogravimetric analysis of ÑBPE. The blue line tracks the percentage of mass remaining given a temperature. The red line is the smoothed inverse of the first derivative of the remaining mass curve. Peaks in the negative first derivative line represent inflection points; temperatures where the mass is changing most quickly. Red numbers depict the temperature at the local maxima of the DTG line. The raw data for this figure is available as Supplemental Dataset 1.

DISCUSSION

Protein content and extraction yield

Raw ÑB has a protein content of about 20% (4). This concentration tends to be on the lower end of other bean and pea cultivars, which tend to have a protein concentration between 20-30% (1,2). ÑB may have a higher starch, moisture, and fiber content, which may relate to its popping properties.

During previous optimization experiments, it was found that both high pH and a large water-to-bean flour ratio improved yield of ÑBPE (12). We decided to use a pH of 10-11 and a ratio of 50 g flour to 750 mL of water for extraction to avoid significant alkaline hydrolysis of the protein and balance experimental constraints. Using lower pH and water-to-bean flour ratios are likely viable routes, but yields may decrease. The typical protein yield for various lots was between 75 and 80%. These yields are reasonable given previously reported isolations using similar conditions on related beans. For instance, isoelectric precipitation using an initial solution step of pH 9.5 yielded 73, 80, 68, and 53 % for chickpea, lentil, broad. and kidney beans (30).

Proximate Analysis

The protein content of ÑBPE was measured using the Biuret, Bradford, and Kjeldahl methods (Table 1); measured concentrations agree closely with each other (<10%). However, there was a 75.4 % recovery of amino acids during the amino acid analysis. This decreased recovery can be explained by incomplete derivatization or hydrolysis, protein glycosylation, or destruction of labile amino acids. The other components measured (fat, ash, moisture, and carbohydrate; Table 1) represent impurities that are likely carryovers from the extraction procedure.

The proximate analysis results of ÑBPE are nearly the same as those for isoelectric precipitates of other beans, such as kidney beans, chickpea, lentil, fava beans, black beans, pea, and soy (1,27, 31-35), where protein concentrations ranged between 70 and 90%. Adjustment of the pH of the acidic precipitation step and a defatting step may also improve the yield and purity of ÑBPE.

Protein Solubility

A U-shaped solubility curve (Figure 1) is characteristic of bean proteins; all tend to have a

pI between 4 and 5, including $\tilde{N}BPE$. Very similar solubility curves have been generated for proteins in cowpeas, kidney bean, peas, lentils, chickpea, common bean, pinto bean, scarlet runner bean, mung bean, and soy (1,2,27,28,33,35,36). This indicates that the isoelectric point and solubility profiles of bean protein extracts are all nearly the same.

Water Absorption

The water absorption of $\tilde{N}BPE$ was measured to be 3.5 g water/ g protein extract. This value is within a factor of two to measurements of other bean protein isolates, such as soy (28), mung bean (36), chickpea (33), fava bean, and pea (34). These differences are based on the solvent accessibility of polar side groups on these protein extracts. It seems that most bean protein isolates, including that of $\tilde{N}B$, absorb water at about 3 ± 2 g water/g protein extract.

Color

$\tilde{N}BPE$ appears as a grayish orange powder with L , a^* and b^* values of 77.34, 0.22, and 16.54, respectively (Table 1). The color of $\tilde{N}BPE$ is very similar to isoelectric bean protein extracts, such as isolates from chickpea, broad bean, and lupin (33,37), kidney beans (32); however, soy protein extract was darker (38).

Viscosity analysis of protein extract

Three measurements of the viscosity of a 25% w/v solution of $\tilde{N}BPE$ yielded a viscosity of 27.5 ± 2 mPa•s (Table 1). A similar analysis completed on great northern bean protein got a nearly identical viscosity of a 10% solution (39). Similarly, work on *P. vulgaris*, and *P. coccineus* proteins indicated a more highly viscous solution than measured here (35). However, this viscosity was similar to lupine bean protein concentrates at a similar concentration (40). These results show that $\tilde{N}BPE$ may be a less efficient viscosity enhancer than other bean protein extracts.

Foaming properties

Measurement and comparison of foaming properties depend on pH, homogenization method, and extract type. Foaming capacity for different protein extracts under different conditions varies widely, from above 500% to 26%; however, the large proportion of bean protein isolates seem to fall

within the range of 50 to 100% foaming capacity (1,2,36,41). The value measured here, 50%, means that half of the foam volume is lost in 20 minutes, which indicates a low-to-moderate foaming capacity. Therefore, the isolate is unsuitable for applications where an abundant stable foam is required; but it may be appropriate where much foaming is not desirable, as in protein drinks.

Electrophoretic analysis

Electrophoretic separation of $\tilde{N}BPE$ (Figure 2) produced three intense bands of mass between 40 and 50 kDa. This electrophoretic pattern is consistent with the subunits of the protein phaseolin. Phaseolins are a major class of bean storage glycoproteins that belong to the 7S vicilin group with a pI of about 5 (1,2,42), also consistent with the pI observed for $\tilde{N}BPE$. It appears that the extraction procedure yielded nearly pure phaseolin with a lack of other common bean proteins, such as lectins and legumins. It has yet to be determined whether the extraction procedure played a role in producing such a pure product, or whether $\tilde{N}B$ has a high phaseolin proportion. The observed banding pattern is remarkably similar to pinto bean, common bean, and scarlet runner bean protein isolate electrophoretic bands, although bands for other proteins are relatively more intense in these samples than in $\tilde{N}BPE$ (25,28,35). Phaseolin easily obtained from $\tilde{N}B$ has relevance as an industrial feedstock, where highly purified phaseolin is desired, such as in weight-loss products, where it has been investigated as diminishing the absorption of carbohydrates by inhibiting the activity of the enzyme α -amylase (43).

Thermogravimetric Analysis

The first temperature inflection point on heating occurred around 60 °C and represented a loss of 5% of the total sample mass. This decrease in mass is most likely the result of water loss from the sample. This percentage loss compares to the measured humidity content (Figure 1). The second inflection occurred around 313 °C and resulted in a loss of about 40% of the mass (Figure 3). This is most likely from the charring of the protein in the $\tilde{N}BPE$. Other smaller inflection points are most likely the result of the pyrolysis of ash and minerals with continued decarbonization of the protein pyrolysis products. The TGA curve observed here is characteristic of protein samples

from pulses such as beans, chickpeas, fava beans, lentils, and peas (44).

Amino acid analysis (AAA)

The AAA results (Table 1) indicate that ÑBPE is particularly rich in glutamine and glutamic acid, asparagine, and aspartic acid, leucine, lysine, phenylalanine, and serine and is deficient in sulfur-containing amino acids. When previously reported amino acid profiles of common pulses, such as pinto (28), common (45, 46), lima (47), tepary (46), soy (28, 31, 48), chickpea (31, 33, 48), lentil (31, 49), pea (50), fava (31, 51), mung (52), African yam (47), mash (53), marama (54) beans and groundnut (48) are compared to each other and the amino acid proportions of ÑBPE, clusters of relative concentration emerge for each amino acid, with all bean AAA profiles following a similar pattern (compare light blue and red dots, Figure 4). In order to quantify this relationship, the amino acid concentrations of ÑBPE were plotted against the amino acid concentrations of the other beans. These comparison plots resulted in almost straight lines, with the coefficient of determination from regression analysis ranging between 0.62 and 0.99, with 85% of the correlations above 0.8 (Supplemental Dataset 2). The sample with the highest correlation to ÑB was a different *P. vulgaris* variety grown in the USA, while Lima beans and tepary beans had the next highest correlations. The samples with the lowest correlation were marama bean and groundnut. Coefficients of determination between all possible combinations of profiles ranged from 0.51 to 1.0; 79% of the coefficients were above 0.8 (Supplemental Dataset 2). Additionally, a two-sided Dixon test on the data in Figure 4 revealed that no ÑB amino acid concentration should be considered an outlier in the data. These data indicate that there is a high correlation between amino acid profiles of ÑB protein extract and other beans. Furthermore, there was a high degree of correlation between the profiles of other bean varieties. This means that all bean varieties, including ÑB, have amino acid profiles that closely resemble each other. This finding holds true over place and time, as Figure 4 compares results obtained over 45 years and includes samples from Africa, Asia, Europe, and North and South America.

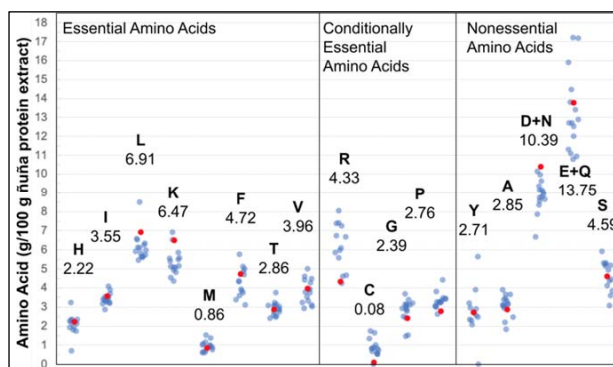


Figure 4. Amino acid profile of ÑBPE (red dots) and its comparison of pinto, common, lima, tepary, soy, lentil, pea, fava, mung, and African yam bean (transparent blue dots). Individual profiles were normalized by dividing the average amino acid content and multiplying by the ÑB average amino acid content. This was necessary because of the different units reported in the literature. Amino acids are represented with one-letter abbreviation, and numbers indicate the amino acid concentration (g/100 g extract) found in ÑBPE. Data and references for this figure are available as Supplemental Dataset 2.

CONCLUSION

From its physical and functional properties, ÑBPE has several nearly identical properties to other isoelectric bean protein isolates and may be well suited for similar food additive applications. Still, it is less efficient at increasing viscosity and forming and maintaining foam. This may work to its advantage because ÑBPE could be used where viscosity and foam are less desirable than nutritional enhancement. Therefore, ÑBPE can enhance the nutritional value of fortified beverages and milk, drinkable or vegan yogurts, weight-loss supplements, or powdered protein supplements for human or animal consumption. It is also possible that using ÑBPE could add unique marketing dimensions to the resulting product, such as connections with Perú and the Andes, organic, non-GMO agriculture, and support of small farmers. Indeed, increasing consumer awareness of other native Andean crops, such as quinoa, amaranth, camu camu, aguaymanto, inka corn, and sacha inchi, has increased demand for these products.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interest with the published results.

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AUTHORS' CONTRIBUTIONS

Tania Vargas: preparation of extract, execution of experiments. Kevin Wilkinson: writing, project conceptualization, data analysis and interpretation, financing of project. Josué Urquiaga: execution of experiments, financing of project. Antonio Rodríguez: conception of project, preparation of extract, planning and execution of experiments, financing of project.

REFERENCES

- Boye J, Zare F, Pletch A. Pulse proteins: processing, characterization, functional properties and applications in food and feed. *Food Res Int.* 2010;43(2):414-431. DOI: <https://doi.org/10.1016/j.foodres.2009.09.003>
- Shevkani K, Singh N, Chen Y, Kaur A, Yu L. Pulse proteins: secondary structure, functionality and applications. *J Food Sci Technol.* 2019;56(6):2787-2798. DOI: <https://doi.org/10.1007/s13197-019-03723-8>
- Valladolid Chiroque AR, Plenge Pardo H, editors. *Leguminosas de Grano: Semillas Nutritivas Para un Futuro Sostenible* [Internet]. Lima, Perú: Ministerio de Agricultura y Riego; 2016 [cited 2019 Nov 25]; p. 42-48. Available from: <https://www.minagri.gob.pe/portal/download/legumbres/catalogo-leguminosas.pdf>
- van Beem J, Kornegay J, Lareo L. Nutritive value of the nuna popping bean. *Econ Bot.* 1992;46(2):164-170. DOI: <https://doi.org/10.1007/BF02930631>
- National Research Council. *Lost Crops of the Incas: Little-known Plants of the Andes with Promise for Worldwide Cultivation* [Internet]. Washington: National Academy Press; 1989 [cited 2019 Nov 25]. 428 p. DOI: <https://doi.org/10.17226/1398>
- Hyland HL. *Plant Inventory No. 172* [Internet]. Washington: U. S. Department of Agriculture; 1968 [cited 2019 Nov 25]. 369 p. Available from: <https://naldc.nal.usda.gov/download/39199/PDF>
- Spaeth SC, Debouck, DG, Tohme J, van Beem J. Microstructure of nunas: Andean popping beans (*Phaseolus vulgaris* L.). *Food Microstruct.* 1989;8(2):263-9. Available from: <https://digitalcommons.usu.edu/cgi/viewcontent.cgi?article=1211&context=foodmicrostructure>
- Zimmer KC. Biological diversity and local development: "popping beans" in the central Andes. *Mtn Res Dev.* 1992;12(1):47-61. <https://doi.org/10.2307/3673747>
- Pesantes MF, Soto JR. Caracterización y evaluación de la variabilidad de doce colectas de *Phaseolus vulgaris* tipo reventon procedente de la provincia de Santiago de Chuco (Perú). *REBIOL.* 2013;33(2):23-33. Available from: <http://revistas.unitru.edu.pe/index.php/faccbiol/article/view/553/516>
- Pearson CH, Ogg JB, Brick MA, Berrada A. Popping and yield characteristics of nuna bean lines developed for temperate climates. *Agron J.* 2012;104(6):1574-8. DOI: <https://doi.org/10.2134/agronj2012.0144>
- Cruz Balarezo J, Camarena Mayta F, Baudoin JP, Huaranga Joaquín A, Blas Sevillano R. Agromorphological evaluation and molecular characterization of popping bean (*Phaseolus vulgaris* L.). IDESEA (Chile). 2009;27(1):29-40. DOI: <http://dx.doi.org/10.4067/S0718-34292009000100005>
- Rodríguez-Zevallos A, Gutiérrez Arce A, Pretel Hiuguay C. Effect of pH on protein extraction and functional properties of Nuna (*Phaseolus vulgaris* L) beans. *Pueblo Continente.* 2014;25(2):81-6. Available from: http://www.upao.edu.pe/publicaciones/PUEBLO_CONTINENTE/pueblo_continente2014_2.pdf
- Castillo Ruiz, R. Caracterización proximal de dos variedades de frijol ñuña (*Phaseolus vulgaris* L.) procedente del distrito de Sarín – provincia de Sánchez Carrión. [Bachelor Thesis]. [Trujillo, Perú]: Universidad Nacional de Trujillo. 2013. 65 p. Available from: <http://dspace.unitru.edu.pe/bitstream/handle/UNITRU/7506/CASTILLO%20RUIZ%20ROSA%20DEL%20PILAR.pdf?sequence=1&isAllowed=y>
- Marmolejo Gutarra KJ. Variabilidad genética del frijol común tipo ñuña (*Phaseolus vulgaris* L.) en las localidades de Carhuaz y Chiquián, Ancash. [Master's Thesis]. [Lima, Perú]: Universidad Nacional Agraria la Molina. 131 p. Available from: <http://repositorio.lamolina.edu.pe/bitstream/handle/UNALM/3398/marmolejo-gutarra-karina-jessica.pdf?sequence=1&isAllowed=y>
- Instituto Nacional de Estadística e Informática, 12.3 Producción de otros productos agrícolas. 2015 (accessed 25 Nov 2019). Available from https://www.inei.gob.pe/media/MenuRecursivo/publicaciones_digitales/Est/Lib1253/cap12/cap12003.xls
- Instituto Nacional de Estadística e Informática. *Compendio estadístico Perú 2018, Capítulo 13: Agrario.* (accessed 25 Nov 2019). Available from: https://www.inei.gob.pe/media/MenuRecursivo/publicaciones_digitales/Est/Lib1635/cap13/cap13.pdf
- Allison, J. WSU professor testing new crops. 2019 (accessed 31 Jul 2020). <https://bit.ly/33aFV21>
- Kisha TJ, Naratto G. Analysis of genetic and nutritional diversity among selected accessions of dry beans and nuna beans (*Phaseolus vulgaris* L.) from the USDA-ARS National Plant Germplasm System. *An. Rept. Bean Improvement Coop.* 2015;58:3-4. Available from: http://arsftfbean.uprm.edu/bic/wp-content/uploads/2018/05/BIC_2015_Annual_Report.pdf
- Cuadros Negri MR, Gómez Carrión J. Performance of "ñuña" seeds *Phaseolus vulgaris* L. (*Fabaceae*) due to the biological fixation of atmospheric nitrogen with *Rhizobium phaseoli*. *Arnaldoa.* 2016;23(1):247-70. Available from: http://cybertesis.unmsm.edu.pe/bitstream/handle/20.500.12672/4871/Cuadros_nm.pdf?sequence=1
- Yuste-Lisbona FJ, Santalla M, Capel C, García-Alcázar M, De La Fuente M, Capel J, De Ron AM, Lozano R. Marker-based linkage map of Andean common bean (*Phaseolus vulgaris* L.) and mapping of QTLs underlying popping ability traits. *BMC Plant Biol.* 2012;12:136. DOI: <https://doi.org/10.1186/1471-2229-12-136>
- González AM, Yuste-Lisbona FJ, Lores M, De Ron AM, Lozano R, Santalla M. Dissecting the genetic basis of popping ability in nuna bean, an ancient cultivar of common bean. *Euphytica.* 2014;196(3):349-63. DOI: <https://doi.org/10.1007/s10681-013-1039-3>
- Ehlers JD, Sterner MH, inventors; Appropriate Engineering and Manufacturing, assignee. Bean-nut popping beans. United States patent 6040503. 2000 Mar 21. Available from: <https://patentimages.storage.googleapis.com/c2/39/0e/a621f152db37c2/US6419976.pdf>
- Chili Smith Family Foods, Nuna. 2019 (accessed 26 Nov 2019). Available from: <https://chilismith.com/product/nuna/>
- Elegant Beans and Beyond, Nuna Bean. (accessed 31 July 2020). Available from: <https://bit.ly/2Ey6TGF>
- Ribeiro, HJS, Prudencio SH, Miyagui DT, Ribeiro EL. Caracterização de concentrado proteico de feijão comum preto, cultivar Iapar 44, novo e envelhecido. *Cienc Tecnol Aliment*

- 2009;29(3):571-80. DOI: <https://doi.org/10.1590/S0101-20612009000300019>
26. Helrich, K. Official Methods of Analysis of the Association of Official Analytical Chemists, 15th ed., Washington: AOAC International; 1990. 1298 p. Available from: <https://law.resource.org/pub/us/cfr/ibr/002/aoac.methods.1.1990.pdf>
27. Jarpa-Parra M, Bamdad F, Wang Y, Tian Z, Temelli F, Han J, Chen L. Optimization of lentil protein extraction and the influence of process pH on protein structure and functionality. *LWT - Food Sci Technol.* 2014;57(2):461-9. DOI: <https://doi.org/10.1016/j.lwt.2014.02.035>
28. Tan ES, Ngoh YY, Gan CY. A comparative study of physicochemical characteristics and functionalities of pinto bean protein isolate (PBPI) against the soybean protein isolate (SPI) after the extraction optimization. *Food Chem.* 2014;152:447-55. DOI: <https://doi.org/10.1016/j.foodchem.2013.12.008>
29. Laemmli UK. Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature.* 1970;227(5259):680-5. DOI: <https://doi.org/10.1038/227680a0>
30. Qayyum MMN, Butt MS, Anjum FM, Nawaz H. Composition analysis of some selected legumes for protein isolates recovery. *J. Anim. Plant Sci.* 2012;22(4):1156-62. Available from: <http://www.thejaps.org.pk/docs/V-22-4/55.pdf>
31. Johnston SP, Nickerson MT, Low NH. The physicochemical properties of legume protein isolates and their ability to stabilize oil-in-water emulsions with and without genipin. *J Food Sci Technol.* 2015; 52(7):4135-45. DOI: <https://doi.org/10.1007/s13197-014-1523-3>
32. Wani IA, Sogi DS, Shivhare US, Gill BS. Physico-chemical and functional properties of native and hydrolyzed kidney bean (*Phaseolus vulgaris* L.) protein isolates. *Food Res Int.* 2015;76(1):11-8. DOI: <https://doi.org/10.1016/j.foodres.2014.08.027>
33. Paredes-Lopez O, Ordorica-Falomir C, Olivares-Vasquez MR. Chickpea protein isolates: physicochemical, functional and nutritional characterization. *J Food Sci.* 1991; 58(3):726-9. DOI: <https://doi.org/10.1111/j.1365-2621.1991.tb05367.x>
34. Fernández-Quintela A, Macarulla MT, Del Barrio AS, Martínez JA. Composition and functional properties of protein isolates obtained from commercial legumes grown in northern Spain. *Plant Foods Hum Nutr.* 1997; 51(4): 331-342. DOI: <https://doi.org/10.1023/A:1007936930354>
35. Makri EA, Doxastakis GI. Emulsifying and foaming properties of *Phaseolus vulgaris* and *coccineus* proteins. *Food Chem.* 2005;98(3):558-68. DOI: <https://doi.org/10.1016/j.foodchem.2005.06.027>
36. Du M, Xie J, Gong B, Xu X, Tang W, Li X, Li C. Extraction, physicochemical characteristics and functional properties of mung bean protein. *Food Hydrocoll.* 2018;76:131-40. DOI: <https://doi.org/10.1016/j.foodhyd.2017.01.003>
37. Alu'datt MH, Rababah T, Alhamad MN, Khalil Ereifej K, Gammoh S, Kubow S, Tawalbeh D. Preparation of mayonnaise from extracted plant protein isolates of chickpea, broad bean and lupin flour: chemical, physicochemical, nutritional and therapeutic properties. *J Food Sci Technol.* 2017;54(6):1395-405. DOI: <https://doi.org/10.1007/s13197-017-2551-6>
38. Zhang QT, Tu ZC, Wang H, Huang XQ, Fan LL, Bao ZY, Xiao H. Functional properties and structure changes of soybean protein isolate after subcritical water treatment. *J Food Sci Technol.* 2015;52(6):3412-21. DOI: <https://doi.org/10.1007/s13197-014-1392-9>
39. Sathe SK, Salunkhe DK. Functional properties of the great northern bean (*Phaseolus vulgaris* L.) proteins: emulsion, foaming, viscosity, and gelation properties. *J Food Sci.* 1981;46(1):71-81. DOI: <https://doi.org/10.1111/j.1365-2621.1981.tb14533.x>
40. Pelgrom PJM, Berghout JAM, van der Goot A, Boom RM, Schutyser MAI. Preparation of functional lupine protein fractions by dry separation. *LWT - Food Sci Technol.* 2014;59(2):680-8. DOI: <https://doi.org/10.1016/j.lwt.2014.06.007>
41. Munc Munc MA, Minka SR, Mbome IL. Optimising functional properties during preparation of cowpea protein concentrate. *Food Chem.* 2014;154:32-7. DOI: <https://doi.org/10.1016/j.foodchem.2013.12.108>
42. De La Fuente M, Lopez-Pedrouso M, Alonso J, Santalla M, De Ron AM, Alvarez G, Zapata, C. In-depth characterization of the phaseolin protein diversity of common bean (*Phaseolus vulgaris* L.) based on two-dimensional electrophoresis and mass spectrometry. *Food Technol Biotechnol.* 2012;50(3):315-25. Available from: https://www.researchgate.net/publication/245032813_In-Depth_Characterization_of_the_Phaseolin_Protein_Diversity_of_Common_Bean_Phaseolus_vulgaris_L_Based_on_Two-Dimensional_Electrophoresis_and_Mass_Spectrometry
43. Yao Y, Hu Y, Zhu Y, Gao, Y, Ren G. Comparisons of phaseolin type and α -amylase inhibitor in common bean (*Phaseolus vulgaris* L.) in China. *Crop J.* 2016; 4(1):68-72. DOI: <https://doi.org/10.1016/j.cj.2015.09.002>
44. Ricci L, Umilta E, Righetti MC, Messina T, Zurlini C, Montanari A, Bronco S, Bertoldo M. On the thermal behavior of protein isolated from different legumes investigated by DSC and TGA. *J Sci Food Agric.* 2018;98(14):5368-77. DOI: <https://doi.org/10.1002/jsfa.9078>
45. Nestares T, Barrionuevo M, Urbano G, López-Frías M. Nutritional assessment of protein from beans (*Phaseolus vulgaris* L) processed at different pH values, in growing rats. *J Sci Food Agric.* 2001;81(15),1522-9. DOI: <https://doi.org/10.1002/jsfa.965>
46. Porch TG, Cichy K, Wang W, Brick M, Beaver JS, Santana-Morant D, Grusak MA. Nutritional composition and cooking characteristics of tepary bean (*Phaseolus acutifolius* Gray) in comparison with common bean (*Phaseolus vulgaris* L.). *Genet Resour Crop Evol.* 2017;64(5):935-53. DOI: <https://doi.org/10.1007/s10722-016-0413-0>
47. Evans MI, Boulter D. Amino acid composition of seed meals of yam bean (*Sphenostylis stenocarpa*) and Lima bean (*Phaseolus lunata*). *J Sci Food Agric.* 1974; 25(8):919-22. DOI: <https://doi.org/10.1002/jsfa.2740250806>
48. Bujang A, Taib NA. Changes on amino acids content in soybean, garbanzo bean and groundnut during pre-treatments and tempe making. *Sains Malays.* 2014;43(4): 503-11. Available from: <https://pdfs.semanticscholar.org/dcc9/9ff030f63b1e7e1d60ed0bdb42b0e5c5854.pdf>
49. Zia-ul-Haq M, Ahmad S, Shad MA, Iqbal S, Qayum M, Ahmad A, Luthria DL, Amarowicz R. Compositional studies of lentil (*Lens culinaris* Medik.) cultivars commonly grown in Pakistan. *Pak J Bot.* 2011; 43(3): 1563-1567. Available from: <https://www.pakbs.org/pjbot/PDFs/43%283%29/PJB43%283%291563.pdf>
50. Leterme P, Monmart T, Baudard E. Amino acid composition of pea (*Pisum sativum*) proteins and protein profile of pea flour. *J Sci Food Agric* 1990;53(1):107-10. DOI: <https://doi.org/10.1002/jsfa.2740530112>
51. Kaldy MS, Kasting R. Amino acid composition and protein quality of eight faba bean cultivars. *Can J Plant Sci.* 1974;54(4):869-71. DOI: <https://doi.org/10.4141/cjps74-155>
52. Shi Z, Yao Y, Zhu Y. Nutritional composition and antioxidant activity of twenty mung bean cultivars in China. *Crop J.* 2016 Oct; 4(5): 398-406. DOI: <https://doi.org/10.1016/j.cj.2016.06.011>
53. Zia-UI-Haq M, Ahmad S, Bukhari SA, Amarowicz R, Ercisli S, Jaafar HZE. Compositional studies and biological activities of some mash bean (*Vigna mungo* (L.) Hepper) cultivars commonly consumed in Pakistan. *Biol Res.* 2014;47(1):23. <https://doi.org/10.1186/0717-6287-47-23>
54. Bower N, Hertel K, Oh J, Storey R. Nutritional evaluation of marama bean (*Tylosema esculentum*, Fabaceae): analysis of the seed. *Econ Bot.* 1988;42(4):533-40. DOI: <https://doi.org/10.1007/BF02862798>